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Title: In vivo stability of WS1321 AASD strains carrying HIV peptide

Date:

Purpose: Determine the number of bacteria recovered from the spleens of orally infected BALB/c mice. Compare the levels of each ^{mutant} strain to that of the parent strain.

Strains: Constructed by Art Branstetter are:

WS1321AASD, PAB102

WS1321AASD, PAB103

WS1321AASD, PAB02::gag

WS1321AASD, PAB03::gag

WS1321AASD, PAB02::vif

WS1321AASD, PAB03::vif

Also

WS1321 - plasmid - Sal. typhimurium strain

See to explain purpose of construction of these 2 strains.

Note: A previous experiment conducted indicated by Day 6 after ^{oral} infection fewer numbers of strain WS1321, PAB103 were recovered from the spleens. ← Lab Notebook #3 (House work)

Experiment: (Full procedure documented in Lab Notebook #3 (House work))

- strains were grown to an O.D. of $\approx 0.8 - 1.0$
- ~~cells~~ to concentrate into DPBS (Some cultures resuspended in 0.35 ml. others into 0.5 ml)
- cultures were diluted & plated to determine ^{amt of} bacteria fed.
- 40ul fed to mice using 200ul pipet.
- mice are to be sacrificed on Days 4, 7 & 11 post-feeding
- 3 mice each strain each timept.

Amt. Given to Mice: Between $3 - 4 \times 10^9$ in 40ul.

Exact #'s are in Book #3

* Spleens homogenized in 2ml HBSS 0.1ml original plated in duplicate

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Results:		ave/splen = ave of 3 x 1/101 x 2nd dil.				
Strain	Day	total ave/splen	Day	total ave/spl.	Day	total ave/spl.
WS1321 #1	4	90, 101	7	128, 122	11	98, 82
#2	131, 46	> 1697	98, 77	> 2097	111, 103	> 1690
#3	70, 71		107, 97		65, 51	
WS1321 #1	44, 37	215	8, 9		63, 100	
PAB102 #2	14, 86	> 1160	92, 135	> 1317	19, 17	> 1123
#3	18, 66		73, 78		74, 67	
WS1321 PAB103 #1	5, 4		35, 46		97, 57	
#2	2, 9	> 70	59, 45	> 767	2, 3	> 1283
#3	0, 0		26, 21		104, 122	
WS1321 PAB102 #1	0, 0		0, 0		2, 4	
#2	0, 0	> 0	18, 19	> 507	0, 0	> 46
#3	0, 0		54, 61		5, 1	
WS1321 #1	0, 0		0, 0		0, 0	
#2	0, 0	> 6	0, 0	> 0	3, 0	> 10
#3	0, 0		0, 0		0, 0	
WS1321 PAB102 #1	0, 0		0, 0		1, 2	
#2	0, 0	> 0	1, 1	> 27	6, 9	> 63
#3	0, 0		0, 6		4, 0	
WS1321 PAB103 #1	0, 0		0, 0		1, 0	
#2	0, 0	> 0	0, 0	> 0	4, 4	> 53
#3	0, 0		0, 0		3, 4	
> 20 not detectable						

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Cloning and Testing HIV genes under the control of the
p_{gag} promoter

Art. Bianchini constructed plasmid pAB104, which
is plasmid pAB103 with the p_{gag} promoter replacing
the lac promoter.

* All data in Notebook #1 Digestions & Ligations
Using directed cloning techniques a 2.7 Kb fragment
encoding VIF from HIV-1 was cloned into pAB104

Buildup:

pAB104 2ul
B buffer 2ul
BamHI 1ul
dH₂O 1ul
20ul

pAB104::VIF 10ul
B buffer 2ul
BamHI 1ul
dH₂O 7ul
20ul

↓ 1 hr. 37°C

- add to each & continue at 37°C
for several hours.

above 20ul
H buffer 3.2ul
SalI 2ul
dH₂O 14.8ul
40ul

- + Each rxn was run on a 1.2% low melt agarose
gel. 1/2 rxn mix on each side of the gel. 1/5 the
gel was stained w/ EtBr after electrophoresis. The
appropriate bands were sliced from the unstained side
- Each band was Prep-A-Mix purified & run
through a Millipore column for purification &
resuspended in 25ul TE for digestion

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Ligation:

Control * PAB104 cut, purified & ligated

10x buffer 5ul

10mM ATP 4ul

PAB104 cut, purified 12.5ul

T4 ligase 2ul

dH₂O 26.5ul

50ul

PAB104 + vif

10x buffer 5ul

10mM ATP 4ul

PAB104 12.5ul

vif 25ul

T4 Ligase 2ul

dH₂O 15ul

50ul

↓ 15°C, overnight in PCR machine

1/2 mix 1/2 mix gel

- Ethanol prec. & electroporate into X6097 ASD

Time (msec)

colonies

Ligation Mix

PAB104 13.7

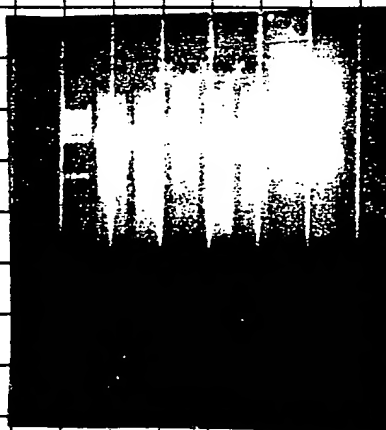
0

PAB104::vif 14.8

many

5 positives were selected for digestion to look for inserts:

- Wizard Miniprep - Purify DNA
- Cut w/ BamH - SmaI
- Run a gel.



all five appear to have inserts.

all five were prepared for Western Blot examination of expression levels. They were electroporated into J6501::ASD for

placement into J6501::ASD

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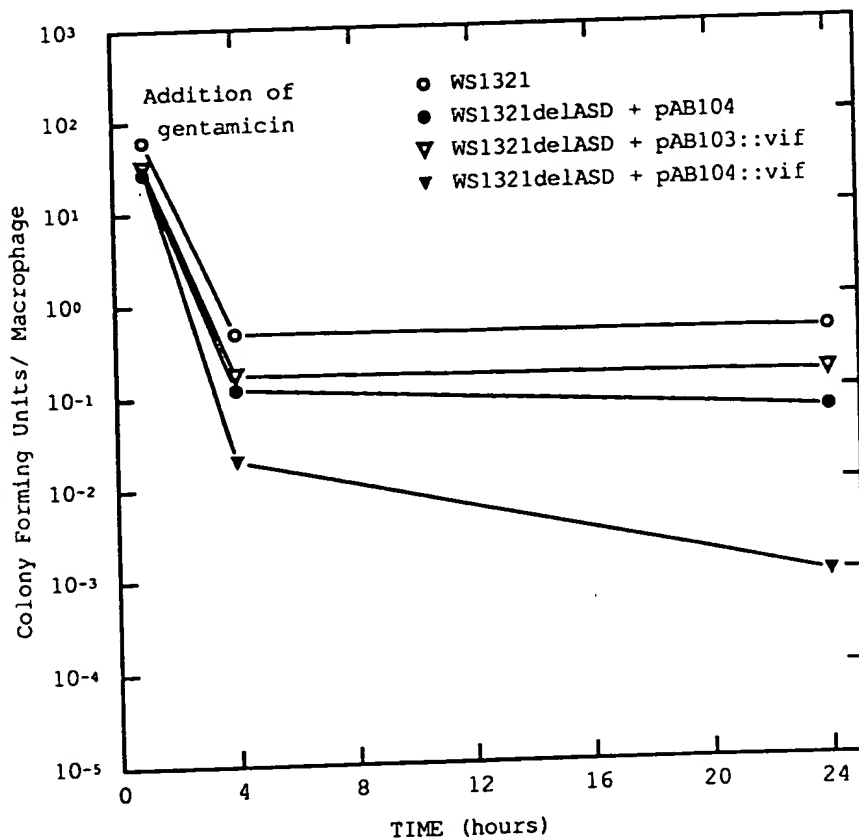
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Title: Macrophage Assay of pagC construct
 Purpose: Determine whether pagC promotes increases
 expression in murine Macrophages

Notebook page #1 full experimental protocol.
 Assay based on protocol of O'Brien et al.
 CFU/Macrophage & Expression via Western blot were assayed.
 Results:



Western: in pagC notebook - Shows expression
 from pagC (pAB104::vif only)
 Slides of Macrophages infected w/ strains - notebook - pagC #1
 Experiment Repeated.

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Title: Growth of WS1321 Δ ASD + pAB104 in Minimal Media
Supplemented w 0.1% Glucose

Notebook page #1

Repeated

Protocol:

- Inoculate colony of each strain into 5ml of LB or MEM.
- Grow 3rd & 4th hrs.
- Concentrate & resuspend in dissociation buffer, vortex freeze & thaw
- Determine protein concentration
- Run Western: compare LB & MEM expression of plac & pagC promoters

Results: (Notebook page #1)

~~3/16~~ EXP.

Bacterial Counts also determined

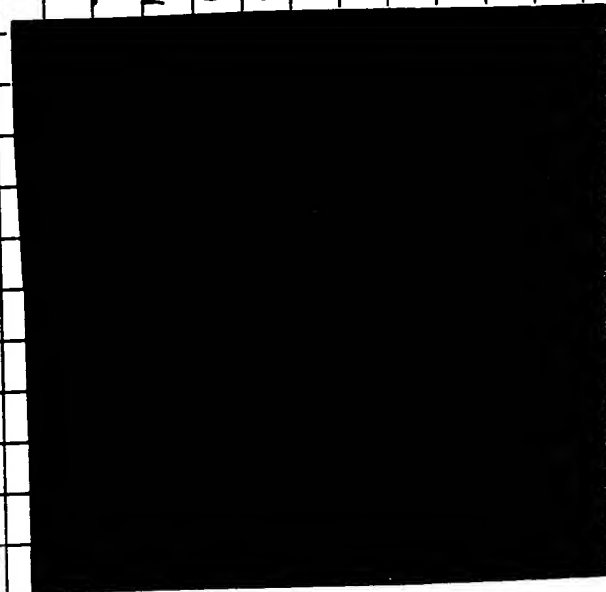
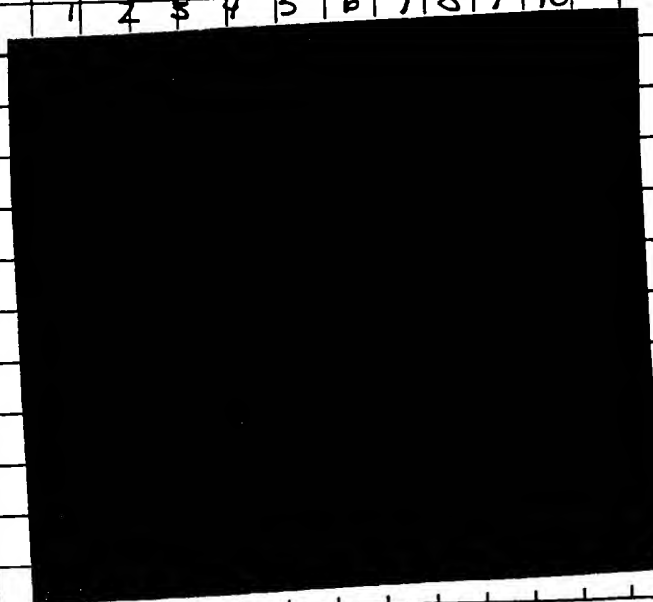
Lane # 10. Marker

LB } 9. WS1321
8 pAB103::vif
7 pAB104
6 pAB104::vif

5 WS1321
4 pAB103::vif
3 pAB104
2 pAB104::vif } MEM

1 2 3 4 5 6 7 8 9 10

1 2 3 4 5 6 7 8 9 10



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Title Mouse Experiment page ConstructsStrains Tested: WS1321WS1321 Δ ASD+ pAB103::vifWS1321 Δ ASD+ pAB104WS1321 Δ ASD+ pAB104::vif

Purpose: Determine the number of bacterial colonies surviving mouse passage & expression of vif gene after passage.
 Mice sacrificed on days 3, 7 & 10

Amount of Bacteria Fed to Mice

40ulWS1321 7.4×10^9 pAB103::vif 4.2×10^9 pAB104 6.9×10^9 pAB104::vif 6.4×10^9

Results: Experimental problem: Apparently mice were mixed by animal handlers. (Based on Western blot analysis)

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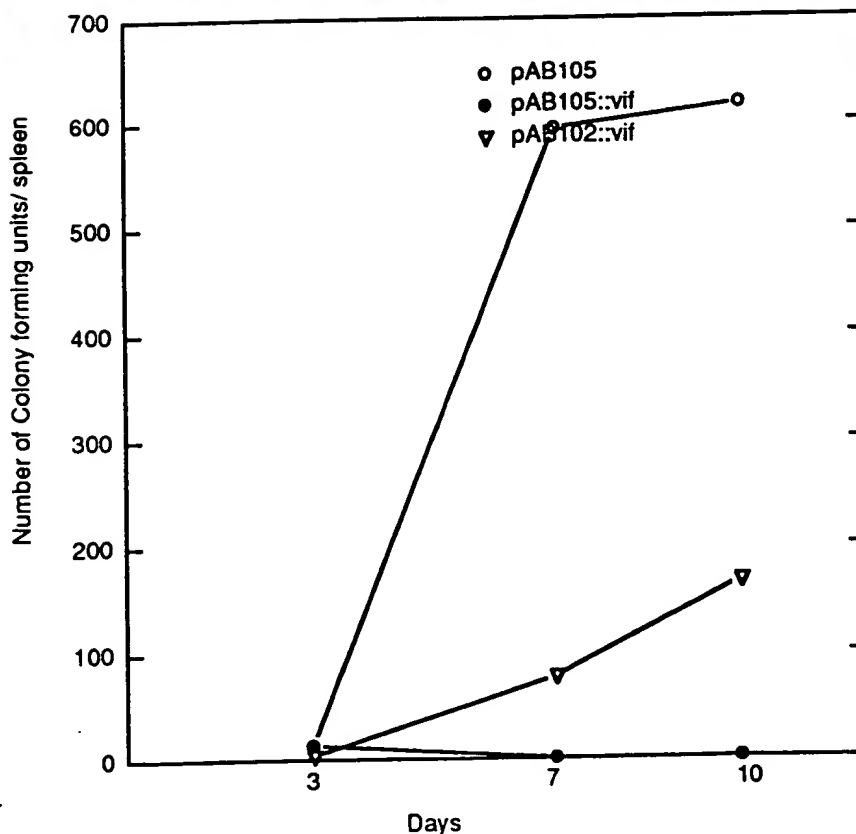
Title: Construction of WS1321ΔASD strains carrying pAB105 & pAB105::vif for mice feeding

Purpose: pAB105 & pAB105::vif plasmids were constructed by OutBranston. In this plasmid the *asd* gene is from *E. coli*. We believe strains carrying plasmids to this *asd* gene are healthier.

5/12 - Mice fed the following strains & amount
46ul

WS1321ΔASD + pAB105	3.6×10^9
WS1321ΔASD + pAB105::vif	2.72×10^9
WS1321ΔASD + pAB102::vif	6.4×10^9

Results: CFU/spleen & expression of vif from recovered colonies



Results of Western
Only pAB102::vif
showed significant
expression.

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Title: Construction of pAB105::ENVITB

Purpose: Of the HIV genes we work with only ENV has mapped CTC epitopes for BALB/c m.c.

After several attempts: → it was determined that the entire ENV gene isn't stable in our pAB105 plasmid. Various deletions resulted after each cloning attempt. Ended July

Will try smaller fragment which has been constructed by Art Blanton

End *Donata R. Seymour*

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Title: Summary of Human Monocyte & S. typhi Interaction Experiments. July

Note: This work has been in progress for some time. This entry is a summary of the work completed & a list of work needed to be done for a publication.

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Trial CTL Assay

Purpose: To standardize non-radioactive CTL assays from Biomega. Testing various Fetal Calf Serum lots.

Testing:

#1 FBS certified (low LDH ≈ 470 units) Gibco
Lot # 40K0242

#2 Hyclone

5ml of #1 or #2
45ml of RPMI w/out Phenol red.
50 ml

- add 50ul to each well
- add 50ul of Control + LDH to 6 wells
- add 10ul of Cyto solution
- add 50ul of Substrate Mix (12ml of Assay Buffer to bottle of Substrate Mix.) Cover plate w foil & incubate at RT for 30'
- Add 50ul of Stop Solution
- Remove large bubbles. Read Ab at 490.

Note - better control RPMI 1640 w/out serum.

Results: #1 best #2 - cannot be used - completely red.
Aug 16 1994

Control Testing Maximin & Spontaneous Release of PB15

#1 Group - RPMI

#2 Group - Media #2 - CTL Assay Media

Note: Cells are healthy. This is a must.

5×10^3 & 1×10^4 cells were tested for spontaneous (maximin) release.

RPMI w/out phenol red & Media #2 w/out phenol red. were used as control backgrounds.

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Results:

	RPMI	Min		Media + 2	
5×10^3	0.479	0.138	$\times 3.5$	0.537	0.153 $\times 3.5$
1×10^4	0.748	0.158	$\times 4.7$	1.083	0.150 $\times 7.22$

$$\text{RPMI} = 0.074$$

$$\text{Media} = 0.162$$

$\rightarrow 3\% \text{ FBS}$

$$\text{Gibco } 10\% = 0.326$$

Assay looks like it should work.

Trial run for CTL Proliferation & Cytokine Assay in *Shigella* 2a, ENVIS-A experiment.

Animals inoculated by Corey Mallet.

Above data contained in *Shigella* Exp/Hela/Hep-2/
Notebook.

Based on info by Art Branstetter a pUC18 plasmid encoding ENVIS-A fragment is stably maintained in *Shigella flexneri* 2a without selection.

Art Branstetter has checked all strains used for this study for Congo Red binding, Toxicity & plasmid content.

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Cloning of ENV_{ITB}-A₁ into pAB105

Purpose: This ENV_{ITB}-A₁ fragment contains the relevant V3 loop, CTL epitope & major Ab epitope. Furthermore, it appears to be more stable in Euc18.

Outline of cloning procedure in rough notes: Page C Notebook

Results: Appears to be cloned. Several weeks of problems to ligase!

Several Ab used for detection of expression

1. Human sera (Auto)
2. Human sera (ITD)
3. MAb R 9-2
4. V3 (ENV_{ITB}-V3-13) - only one that is good!

Gels Run

Comparing pAB102::ENV_{ITB}-A₁ (plac) to pAB105::ENV_{ITB}-A₁ (page) +
 Oug/well protein

Repeated Results: - plac lysates from Art for 11-28-94 not enough protein loaded

- page expression is clearly higher than plac.

Need to repeat w/ fresh Ab, but all looks good.
 Set-up for mouse testing.

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Shigella CMI Response50 mice total: 4th floor

Purpose: Test for CTL activity to EN1 epitope.

Mice sacrificed Sept. 6: Note: animals look ruffled.
- 2 *S. flexneri* 2a died.

Cells prepared for CTL, proliferation & cytotoxic assays.

Concept dropped

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Previous background experiments appear in
Notebooks 1 & 2.

As of , to extend this concept we
will try antibiotic treatment of BHK cells which
have been infected w/ SC602(pCMVp). The
idea is that antibiotic treatment will kill
the intracellular bacteria. The dead bacteria
will release the pCMVp plasmid into the cytoplasm
for transcription & translation by the BHK cell.

This entry is based on a conversation with
Jung Hwang on .
made on .

- This entry was
written by Donna R. Sigmund

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